

# Prevalence and Genotype of *Giardia duodenalis* of Asymptomatic Individual in a Child Care Center, Bangkok, Thailand

Duangnate Pipatsatitpong<sup>1\*</sup>, Ratchaneewan Aunpad<sup>1</sup>

<sup>1</sup>Faculty of Allied Health Sciences, Thammasat University, Pathum Thani 12120, Thailand

Suradej Siripattanapipong<sup>2</sup>

<sup>2</sup>Department of Microbiology, Faculty of Science, Mahidol University, Bangkok 10400, Thailand

Mathirut Mungthin<sup>3</sup>, Saovanee Leelayoova<sup>3</sup>

<sup>3</sup>Department of Parasitology, Phramongkutklao College of Medicine, Bangkok 10400, Thailand

Received 11 April 2018; Received in revised form 31 July 2018

Accepted 27 August 2018; Available online 25 September 2018

## ABSTRACT

*Giardia duodenalis* (*G. duodenalis*) is an intestinal flagellate protozoan, which infects in humans and animals. It can be causing giardiasis. The symptoms are acute or chronic diarrhea, abdominal pain, weight loss and dehydration. *G. duodenalis* can be transmitted by eating food or drinking water contaminated with infected stage cysts. Children who live and share facility in an orphanage are the high risk groups for infection. The objective of this paper is to study the prevalence and genotypes of *G. duodenalis* infection of asymptomatic individual in a child care center. This study was conducted in Mercy Centre orphanage, Klong Toey community. 289 stool samples were collected from children and childcare workers. Stool samples were extracted DNA. Small subunit ribosomal DNA (SSU-*rDNA*) and glutamate dehydrogenase (*gdh*) genes were amplified using Nested PCR. PCR products of *gdh* gene were cut using *Nla*IV and *Rsa*I restriction enzymes. Genotypes were identified. The results of this study showed the prevalence of giardiasis in an orphanage, Klong Toey community, Thailand was 17.6% (51/289). PCR-RFLP analysis of 19 samples revealed that genotype AII was 63.2% (12/19), genotype BIV was 31.6% (6/19) and genotype BIII was 5.3% (1/19) respectively. Consequently, the study of prevalence and genotypes of *G. duodenalis* is beneficial for control planning of giardiasis in an orphanage, Klong Toey community.

**Keywords:** *Giardia duodenalis*; Genotype; Assemblage

## 1. Introduction

*Giardia duodenalis* (*G. duodenalis*) is an intestinal flagellate protozoan, causing giardiasis, which infects a wide range of host, such as humans, domestic animals. *G. duodenalis* is one of the most intestinal protozoan that distributes worldwide [1-3]. The prevalence of *G. duodenalis* is found in all age and children are the greatest risk group for infection. The prevalences of *G. duodenalis* infection range from 1.3% to 37.7% in children, Thailand [4-7]. Infection occurs by ingestion of viable cysts, which are transmitted by fecal-oral route, person to person or animal to person, water borne and food borne. The symptoms produce acute or chronic diarrhea, malabsorption, weight loss and abnormal growth in children [1-3]. *G. duodenalis* has eight major genotypes, genotype A-H. Genotype A and B are associated with humans. Other animal-specific groups include genotype C, D, E, F and G. Genotype A has two distinct clusters, AI and AII, while genotype B consists of BIII and BIV [1-3]. The genotypes infected in Thailand are sub-genotype such as BIV, AII, BIII and AI respectively. Nowadays, the common identify method of *G. duodenalis* is microscopic method in direct stool smear. However, microscopic method is high specificity and easily to identify but it has low sensitivity. Immunofluorescent method is superior sensitivity but it is also more expensive. These techniques might not be sensitive enough to detect low number of *Giardia* cysts and cannot be used to discriminate genotype of *G. duodenalis*. Therefore, nested polymerase chain reaction (Nested PCR) method has been developed. It uses 2 sets of primer that aims to increase specificity and number of PCR products. Nested PCR method uses for diagnosis

which is higher sensitivity than microscopic method and immunofluorescent method.

Because it can detect a low number of *Giardia* cysts [3]. PCR-RFLP technique uses for determine genotype and sub-genotype of *G. duodenalis*. Furthermore, PCR-RFLP can be clearing to analyze mix genotypes. In Thailand, many study of *G. duodenalis* were conducted only in some especially rural community such as Chacheongsao province and Mae-chaem, Hod districts, Chiang Mai province [5, 7]. At the present time, information about prevalence and genotype of giardiasis in urban community of Thailand are not available. Consequently, the objectives of this study were determined the prevalence and the genotype of *G. duodenalis* in an orphanage, Klong Toey community.

## 2. Materials and Methods

### 2.1 Stool collection

Two hundred and eighty nine stool specimens were collected from children and child care workers in the Mercy Centre orphanage, Klong Toey community during September 2009 to September 2010, participants were voluntarily enrolled into the previous study, with the written informed consent from their parents or a head of child care institute. Personal data of children was provided by their parents or child care workers. All stool specimens were kept at -20°C without preservative agent. Research protocol in this study was approved by the Ethics Committee of Thammasat University.

### 2.2 DNA extraction

All stool samples were extracted DNA using QIAmp DNA stool mini kit (QIAGEN, Germany) according to manufacturer instructions [8].

### 2.3 Genotypic characterization of *Giardia duodenalis*

Genotypic characterization of *G. duodenalis* was determined using amplification of the small subunit ribosomal

gene (SSU-*rRNA*) as previously described by Hopkins RM, et al. [9].

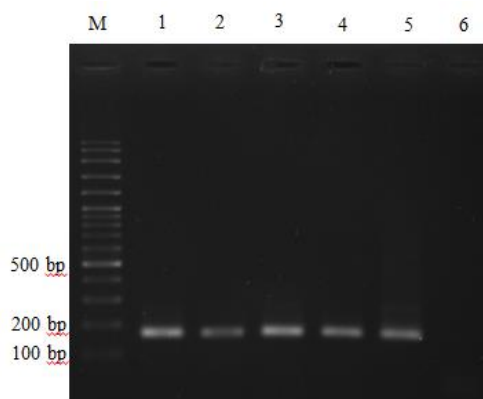
PCR-RFLP was also performed using nested PCR of 461 bp region of the glutamate dehydrogenase (*gdh*) gene. The primary primer pairs (forward primers; GDH1, GDH1a and reverse primer; GDH5s) with the condition for the first round PCR were used as previously described by Boontanom P, et al., 2011 [5]. The secondary PCR, forward primer GDHeF and reverse primer GDHiR were used and the condition was described by Read CM, et al. 2004 [3]. To identified genotype and sub-genotype of *G. duodenalis*, a 461 bp of the *gdh* gene was digested with restriction enzyme *NlaIV* to differentiate all major genotype including sub-genotype AI and AII. A restriction enzyme *RsaI* was used to distinguish between sub-genotype BIII and BIV. PCR products and restriction fragments were separated by electrophoresis in 1.5% agarose gel. Gels were stained with ethidium bromide and visualized under UV light and documented by using a gel documentation system. All positive PCR product samples were confirmed by DNA sequencing (AIT Biotech Company). The genotype of *G. duodenalis* from each specimen was confirmed based on the homology of the sequenced PCR product to the published sequence in GenBank.

### 3. Results and Discussion

#### 3.1 Results

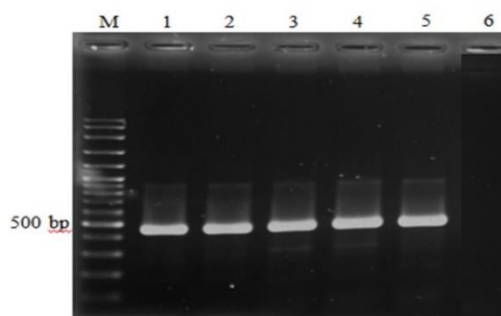
A total of 289 stool samples from an orphanage, Klong Toey community Thailand were analyzed by nested PCR using amplification of the SSU-*rDNA* gene and *gdh* gene. Using RH11/RH4 as primary primers and GiarF/GiarR as secondary primers for SSU-*rDNA* gene (Figure 1), 51 samples (17.6%) were positive results. Using *GDH1/GDH1a/GDH5s* as primary primers and *GDHeF/GDHiR* as secondary primers for *gdh* gene (Figure 2), 19 samples (6.6%) were positive results. For diagnostic

genotyping, *gdh* PCR products could be used RFLP analysis by *NlaIV* restriction enzyme for AI, AII genotype identifications and *RsaI* restriction enzyme for identified BIII and BIV genotypes (Figure 3-4). Of 19 samples, 12 samples (63.2%) were classified into genotype AII, 1 sample (5.3%) was genotype BIII, 6 samples (31.6%) were genotype BIV, whereas genotype AI was not found.



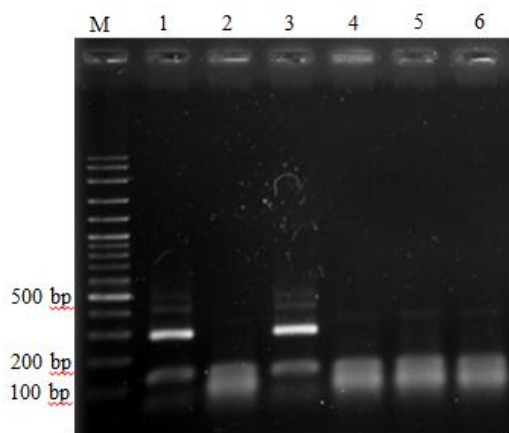
**Fig.1.** Nested PCR amplification of *Giardia duodenalis* SSU-*rDNA* gene.

Lane M; 100 bp markers, lane 1-4; stool samples, lane 5; positive control, lane 6; negative control.



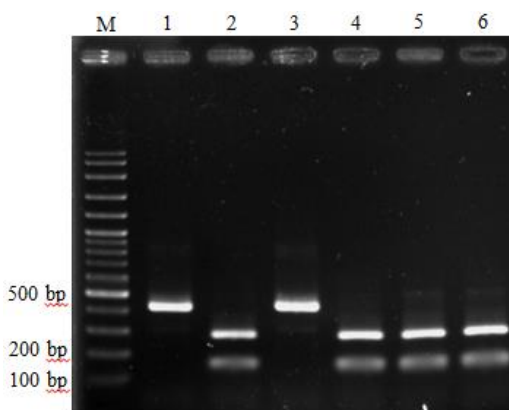
**Fig.2.** Nested PCR amplification of *G. duodenalis*; *gdh* gene.

Lane M; 100bp markers, lane 1-4; stool samples; lane 5; positive control, lane 6; negative control.



**Fig.3.** RFLP analysis using *NlaIV* restriction enzyme.

Lane M; 100 bp markers, lane 1 and 3; Suspected B genotype, lane 2, 4-6; AII genotype.



**Fig.4.** RFLP analysis using *RsaI* restriction enzyme.

Lane M; 100 bp markers, lane 1 and 3; BIV genotype, lane 2, 4-6; BIII genotype.

### 3.2 Discussion

The prevalences of giardiasis of the children population in Thailand were previously reported in many studies. The prevalence of giardiasis in children varies from 1.3 to 37.7%. [4-7]. In 2008, a study in primary school of a rural community, Chacheongsao province showed a prevalence of 6.2% [6]. Next, in 2011, a study in preschool at Sanamchaiket District, Chacheongsao province showed a

prevalence of 5.8% [5]. The last, in 2012, a study in hilltribe children, Mae-chaem and Hod districts, Chiang Mai showed a prevalence of 5.2% [9]. However, the present study showed the prevalence of orphanage was 17.6%. The high prevalence not only depended on sanitary living place and their hygiene but also depended on sensitivity of the identify methods, Molecular technique such as conventional PCR, nested PCR have higher sensitivity than microscopic method.

The genotypes of *G. duodenalis* infection in humans were genotype A (AI, AII) and B (BIII, BIV). In 2011, a study in preschool children at Sanamchaiket District, Chacheongsao province showed the most frequent genotype of *G. duodenalis* was BIV, followed by AII and BIII respectively whereas genotype AI was not found [5]. This result was the same as the study in primary schoolchildren of a rural community, Chacheongsao province [6] and the study in hilltribe children, Hod district, Chiang Mai [7]. The study in hilltribe children, Mae-chaem district, Chiang Mai showed the most frequent genotype was AII, followed by BIV whereas genotype AI and BIII were not found. However, this study showed the most frequent genotype was AII, followed by BIV and BIII respectively whereas genotype AI was not found.

In 2014, Boontanom P, et al. reported the genotypes of *Giardia. duodenalis* from stool sample freezing. Stool samples were collected from children in an orphanage in suburban area outside Bangkok, Central Thailand during January 2007 to January 2008 [10]. Genotype AII was the most predominant found, followed by genotype BIV in an orphanage cohort.

During January 2015 to August 2018 [11-13], many studies investigated parasite infections among children in Thailand [11-13], *G. duodenalis* was still a highlight protozoan infection among children.

However, there have no report of genotypes determination.

Comparison of the diagnostic tests; such as microscopy, ELISA and PCR for the detection of *G. duodenalis*. The sensitivity of the PCR technique was higher than the ELISA and microscopy technique [14]. PCR-RFLP technique can be used to discriminate genotype of *G. duodenalis*.

#### 4. Conclusion

The prevalence of giardiasis in an orphanage, Klong Toey community was 17.6%. PCR-RFLP analysis of 19 stool samples revealed that genotype AII (63.2%), BIV (31.6%) and BIII (5.3%) were found, whereas genotype AI was not found. This information would be help to develop a treatment planning, preventing and controlling of giardiasis in an orphanage, Klong Toey community, Thailand.

#### Acknowledgements

The authors would like to thank all participants, who enrolled in this study. For conflict of interests, the authors declare that they have no conflicts of interest in this study.

This study was financially supported by research grants from Thammasat University.

#### References

- [1] Monis, PT, Andrews RH, Mayrhofer G, Ey PL. Genetic diversity within the morphological species *Giardia intestinalis* and its relationship to host origin. *Infect Genet Evol* 2003; 3(1): 29-38.
- [2] Plutzer J, Ongerth J, Karanis P. *Giardia* taxonomy, phylogeny and epidemiology: Facts and open questions. *Int J Hyg Environ Health*. 2010; 213(5): 321-33.
- [3] Read CM, Monis PT, Thompson RC. Discrimination of all genotypes of *Giardia duodenalis* at the glutamate dehydrogenase locus using PCR-RFLP. *Infect Genet Evol* 2004; 4(2): 125-30.
- [4] Pipatsatitpong D, Leelayoova S, Mungthin M, Aunpad R, Naaglor T, Rangsin R. Prevalence and risk factors for *Blastocystis* infection among children and caregivers in a child care center, Bangkok, Thailand. *Am J Trop Med Hyg* 2015; 93(2): 310-5.
- [5] Boontanom P, Mungthin M, Tan-Ariya P, Naaglor T, Leelayoova S. Epidemiology of giardiasis and genotypic characterization of *Giardia duodenalis* in preschool children of a rural community, central Thailand. *Trop Biomed* 2011; 28(1): 32-39.
- [6] Ratanapo S, Mungthin M, Soontrapa S, Faithed C, Siripattanapipong S, Rangsin R, et al. Multiple modes of transmission of giardiasis in primary schoolchildren of a rural community, Thailand. *Am J Trop Med Hyg* 2008; 78(4): 611-5.
- [7] Saksirisampant W, Boontanom P, Mungthin M, Tan-Ariya P, Lamchuan D, Siripattanapipong S, et al. Prevalence of giardiasis and genotypic characterization of *Giardia duodenalis* in hilltribe children, Northern Thailand. *Trop Biomed* 2012; 29(3): 331-338.
- [8] QAIGEN [online] Available from: <https://www.qiagen.com/us/shop/sample-technologies/dna/genomic-dna/qiaamp-fast-dna-stool-mini-kit/#orderinginformation>. Accessed April 11, 2018.
- [9] Hopkins RM, Meloni BP, Groth DM, Wetherall JD, Reynoldson JA, Thompson RC. Ribosomal RNA sequencing reveals differences between the genotypes of *Giardia* isolates recovered from humans and dogs living in the same locality. *J Parasitol* 1997; 83(1): 44-51.
- [10] Boontanom P, Pipatsatitpong D, Tan-ariya P, Mungthin M, Siripattanapipong S, Naaglor T, Leelayoova, S. Incidence and risk factors of *Giardia duodenalis* infection in an orphanage, Thailand. *Trop Biomed* 2014; 31(3): 525-33.
- [11] Assavapongpaiboon B, Bunkasem U, Sanprasert V, Nuchprayoon S. A cross-sectional study on intestinal parasitic infections in children in suburban public primary schools, Saraburi, the central region of Thailand. *Am J Trop Med Hyg* 2018; 98(3): 763-67.

- [12] Yanola J, Nachaiwieng W, Duangmano S, Prasannarong M, Somboon P, Pornprasert S.  
Current prevalence of intestinal parasitic infections and their impact on hematological and nutritional status among Karen hill tribe children in Omkoi District, Chiang Mai Province, Thailand. *Acta Trop* 2018; 180: 1-6.
- [13] Sanprasert V, Srichaipon N, Bunkasem U, Srirungruang S, Nuchprayoon S.  
Prevalence of intestinal protozoan infections among children in Thailand: A large-scale screening and comparative study of three standard detection methods. *Southeast Asian J Trop Med Public Health* 2016; 47(6): 1123-33.
- [14] Van den Bossche D, Cnops L, Verschueren J, Van Esbroeck M. Comparison of four rapid diagnostic tests, ELISA, microscopy and PCR for the detection of *Giardia lamblia*, *Cryptosporidium* spp. and *Entamoeba histolytica* in feces. *J Microbiol Methods* 2015; 110: 78-84.