



Received 11<sup>th</sup> September 2020,  
Revised 06<sup>th</sup> October 2020,  
Accepted 06<sup>th</sup> October 2020

[DOI: 10.14456/past.2020.6](https://doi.org/10.14456/past.2020.6)

## Xanthones and Coumarins from the Twigs of *Mesua ferrea* L.

Suda Chakthong<sup>1,2,3\*</sup>, Arnon Chukaew<sup>1,2</sup>, Saowanit Saithong<sup>1,3</sup>, Sasitorn Chusri<sup>2,4</sup>, Surasak Limsuwan<sup>2,4</sup> and Supayang P. Voravuthikunchai<sup>2,5</sup>

<sup>1</sup>Division of Physical Science, Faculty of Science, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand.

<sup>2</sup>Natural Product Research Center of Excellence, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand.

<sup>3</sup>Center of Excellence for Innovation in Chemistry, Faculty of Science, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand

<sup>4</sup>Faculty of Traditional Thai Medicine, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand.

<sup>5</sup>Division of Biological Science, Faculty of Science, Prince of Songkla University, Hat Yai, Songkhla, 90112, Thailand.

\*E-mail: suda.ch@psu.ac.th

### Abstract

Five xanthones **1–5** and four phenylcoumarins **6–9** were isolated from acetone extracts of twigs of *Mesua ferrea* L. The structures of these compounds were elucidated by spectroscopic analysis and comparison with reported data such as 1D and 2D NMR, UV and IR spectroscopy. Compounds **4–7** were isolated from this plant for the first time. Compound **3**, 1,5,6-trihydroxyxanthone, showed significant antioxidant activities with IC<sub>50</sub> values of free radical scavenging in DPPH and ABTS assays of 45.0 and 197.0 µg/mL, respectively, as compared to the control, trolox, (77.6±1.0 µg/mL and 282.9±1.2 µg/mL for DPPH and ABTS assay, respectively). Besides, mammea A/AA cyclo F, compound **9**, formed in a single crystal and was reported.

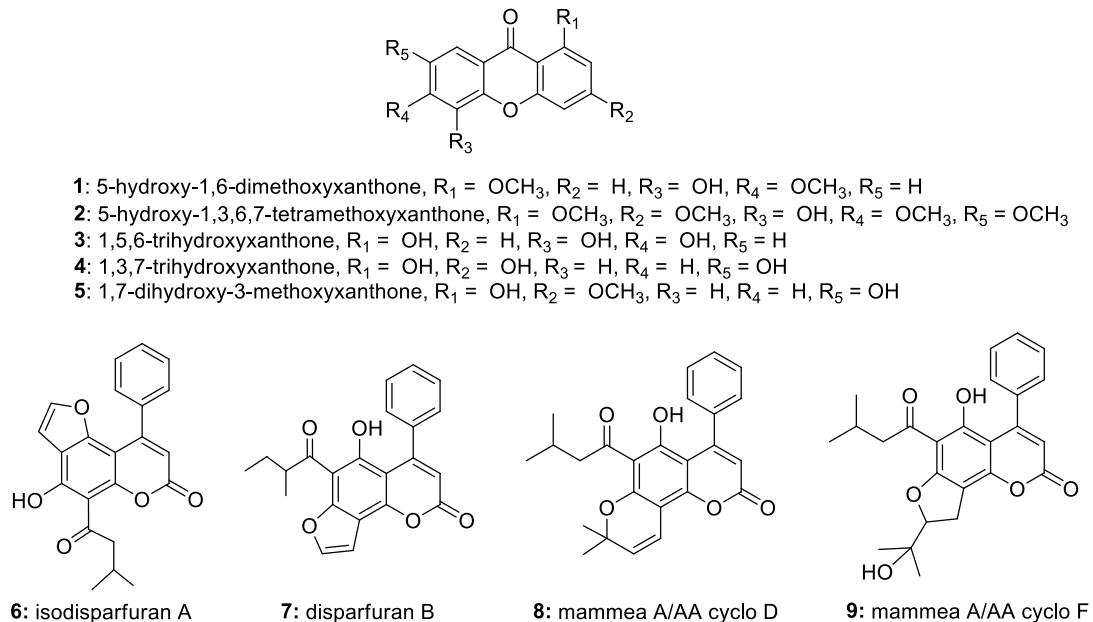
**Keywords:** *Mesua ferrea* L., Calophyllaceae, xanthones, phenylcoumarins

### 1. Introduction

*Mesua ferrea* L., is known locally as “bunnak”, belongs to the Calophyllaceae family and is distributed throughout Southeast Asia, India and Sri Lanka. In India, aerial parts of this plant have been used in traditional medicine to treat various diseases. A decoction of the flowers with sugar candy is used to stop bloody stool (1). The essential oil of fruits and flowers was used to treat rheumatism and cure skin diseases (2). The dried flowers exhibit stomachic properties and anti-inflammatory (3). The seed oil is a safe pharmaceutical excipient and could be used in medicinal formulations (4). *M. ferrea* is used as an anticancer, an antimicrobial, an antipyretic, a cardiotonic, a carminative, a diuretic and an expectorant agent (5, 6). In Thailand, the bark is traditionally used for the treatment of cough, dysentery, and vomiting (7), while the seeds are used as an aromatic, an expectorant, and a wound healer (8).

We previously reported the isolation of several xanthones from roots of *M. ferrea* and some of those compounds had cytotoxic properties (9). In

this study nine known compounds, including five xanthones **1–5** and four phenylcoumarins **6–9**, are isolated and characterised from twigs of *M. ferrea* (Figure 1), identified as 5-hydroxy-1,6-dimethoxyxanthone **1** (9), 5-hydroxy-1,3,6,7-tetramethoxyxanthone **2** (10), 1,5,6-trihydroxyxanthone **3** (11), 1,3,7-trihydroxyxanthone **4**, 1,7-dihydroxy-3-methoxy xanthone **5** (12), isodisparfuran A **6**, disparfuran B **7** (13), mammea A/AA cyclo D **8** (14) and mammea A/AA cyclo F **9** (15, 16). Although these compounds are known compounds, to the best of our knowledge no NMR data have been published for compounds **1** and **2**. Therefore, the present report presents the complete assignment of <sup>1</sup>H and <sup>13</sup>C-NMR spectral data of compounds **1** and **2** along with the single-crystal X-ray diffraction analysis of compound **9**. Also, compounds **4–7** are isolated from this plant and reported for the first time. Compounds **3–5** and **9** are evaluated for their antioxidant properties based on the DPPH and ABTS radical scavenging activities and antibacterial activity.



**Figure 1** Structures of compounds **1–9**

## 2. Materials and Experiment

### 2.1 Plant material

The twigs of *Mesua ferrea* L. were collected from Trang Province, Thailand, in April 2017 and the identification was made by Mr. Sukid Ruangrua, the Forest Herbarium (BKF). A voucher specimen (BKF NO. 194350) has been deposited at the Forest Herbarium (BKF), Chatuchak, Bangkok.

### 2.2 General experimental procedures

The infrared (IR) spectra were recorded on a PerkinElmer 783 FTS 165 FT-IR spectrometer. Ultraviolet (UV) absorption spectra were measured in MeOH on a UV-160A spectrophotometer (Shimadzu). The <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on 300 MHz and 500 MHz Bruker FTNMR Ultra Shield spectrometer with tetramethylsilane (TMS) as an internal standard. Quick column chromatography (QCC) was carried out on silica gel 60 GF<sub>254</sub> (Merck). Column chromatography (CC) was performed using silica gel (Merck) type 100 (70–230 mesh ASTM). Thin-layer chromatography (TLC) and precoated TLC (PTLC) were carried out on silica gel 60 F<sub>254</sub> (Merck).

### 2.3 Extraction and isolation of compounds

The acetone extract was extracted for one week from 8.3 kg of the twigs with acetone at room temperature. A brownish crude acetone extract was furnished at a mass of 43.0 g which was subjected to QCC using a hexane eluent. Polarity was increased with acetone and MeOH. The eluates were combined based on TLC characteristics to afford 13 fractions (F1–F13). Fraction F5 was purified by CC with acetone–hexane (1.5:4.0, v/v) to yield **4** (12.5 mg)

and **7** (2.1 mg). Fraction F6 gave **5** (3.1 mg). Compounds **8** (11.5 mg) and **9** (10.5 mg) were isolated from fraction F10 (615.0 mg). Fractions F11 (6.4 g) was purified by QCC with a gradient of hexane–CH<sub>2</sub>Cl<sub>2</sub>–EtOAc to yield **3** (2.1 mg) and **6** (1.3 mg). Compounds **1** (4.3 mg) and **2** (1.5 mg) were isolated from fractions F13 (5.6 mg) and F12 (145.5 mg), respectively.

#### 2.3.1 5-hydroxy-1,6-dimethoxyxanthone (**1**)

Pale yellow solid, UV  $\lambda_{\text{max}}$  (MeOH) (log  $\varepsilon$ ): 240 (3.95), 263 (3.42) and 344 (4.17); IR (neat)  $\nu_{\text{max}}$ : 3280, 1650 and 1575 cm<sup>-1</sup>. <sup>1</sup>H NMR (acetone-*d*<sub>6</sub>, 300 MHz)  $\delta$ : 3.94 (3H, s, 1-OCH<sub>3</sub>), 3.98 (3H, s, 6-OCH<sub>3</sub>), 6.94 (1H, d, *J* = 8.0 Hz, H-2), 7.10 (1H, d, *J* = 8.7 Hz, H-7), 7.11 (1H, d, *J* = 8.0 Hz, H-4), 7.67 (1H, d, *J* = 8.7 Hz, H-8), 7.67 (1H, t, *J* = 8.0 Hz, H-3). <sup>13</sup>C NMR (acetone-*d*<sub>6</sub>, 75 MHz)  $\delta$ : 55.6 (1-OCH<sub>3</sub>), 55.9 (6-OCH<sub>3</sub>), 106.0 (C-2), 108.2 (C-7), 109.7 (C-4), 112.0 (C-9a), 116.4 (C-8), 117.9 (C-8a), 134.0 (C-5), 134.6 (C-3), 144.3 (C-10a), 151.6 (C-6), 158.2 (C-4a), 160.9 (C-1), 174.3 (C-9).

#### 2.3.2 5-hydroxy-1,3,6,7-tetramethoxyxanthone (**2**)

Pale yellow solid; UV  $\lambda_{\text{max}}$  (MeOH) (log  $\varepsilon$ ): 215 (3.61), 263 (3.03) 317 (3.17) and 382 (3.82); IR (neat)  $\nu_{\text{max}}$ : 3125, 1623 and 1578 cm<sup>-1</sup>. <sup>1</sup>H NMR (CD<sub>3</sub>OD, 500 MHz)  $\delta$ : 3.91 (3H, s, 3-OCH<sub>3</sub>), 3.92 (3H, s, 1-OCH<sub>3</sub>), 3.93 (3H, s, 6-OCH<sub>3</sub>), 3.94 (3H, s, 7-OCH<sub>3</sub>), 6.48 (1H, d, *J* = 2.0 Hz, H-2), 6.74 (1H, d, *J* = 2.0 Hz, H-4), 7.13 (1H, s, H-8). <sup>13</sup>C NMR (CD<sub>3</sub>OD, 125 MHz)  $\delta$ : 55.0 (7-OCH<sub>3</sub>), 55.1 (6-OCH<sub>3</sub>), 55.2 (1-OCH<sub>3</sub>), 59.8 (3-OCH<sub>3</sub>), 92.7 (C-4), 94.7 (C-8), 94.8 (C-2), 106.0 (C-9a), 118.0 (C-8a),

137.3 (C-5), 142.3 (C-6), 150.2 (C-7), 150.4 (C-10a), 160.1 (C-4a), 161.7 (C-1), 165.3 (C-3), 178.8 (C-9).

#### 2.4 X-ray crystallographic of **9**

The crystal data of **9** were collected on CCD SMART APEX diffractometer with graphite-monochromated Mo K $\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ) at 296(2) K with 13086 reflections. The SMART program was utilized for data collection and SAINT Program was used for cell refinement and data reductions, respectively (17). Absorption correction was applied by using the multi-scan program SADABS (17). The structure was solved by direct methods refined by a full-matrix least-squares procedure based on  $F^2$  with the SHELXT program packages (18). The C-bound H atoms were positioned geometrically with the distance of C—H = 0.93  $\text{\AA}$ ,  $U_{iso} = 1.2U_{eq}$  for C- $sp^2$  (for aromatic H atoms) and distances of C—H = 0.96–0.98  $\text{\AA}$ ,  $U_{iso} = 1.5U_{eq}$  for C- $sp^3$ , respectively. All H atoms of carbon atoms are constrained to ride on their parent atoms. The hydrogen atoms of hydroxyl groups and water molecules were located in a difference map and the coordinates were refined isotropically and the O—H distances are in the range of 0.79(5)–0.90(4)  $\text{\AA}$ . The WinGXv2014.1 and Mercury programs were used to prepare the materials and molecular graphic for publication (18, 19, 20). Crystallographic data for **9** were deposited at the Cambridge Crystallographic Data Center (CCDC Number: 1998635). Copies of the data can be obtained, free of charge, through application to the Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK, (fax: +44-(0)1223-336033, or by e-mail at <http://www.ccdc.cam.ac.uk/deposit@ccdc.cam.ac.uk>).

#### 2.4.1 Crystallographic data of *Mammea A/AA cyclo F* (**9**)

Colourless crystals,  $C_{25}H_{26}O_6 \cdot H_2O$ ,  $M = 440.47$ ,  $W = 0.71073 \text{ \AA}$ ,  $0.202 \times 0.156 \times 0.104 \text{ mm}^3$ , monoclinic,  $P2_1/c$ ,  $a = 9.7967(5) \text{ \AA}$ ,  $b = 21.7164(10) \text{ \AA}$ ,  $c = 11.3173(6) \text{ \AA}$ ,  $\alpha = 90.00^\circ$ ,  $\beta = 110.0020(10)^\circ$ ,  $\gamma = 90.00^\circ$ ,  $V = 2262.5(2) \text{ \AA}^3$ ,  $Z = 4$ ,  $D_x = 1.293 \text{ Mg/m}^3$ , crystal size  $0.202 \times 0.156 \times 0.104 \text{ mm}^3$ ,  $\mu(\text{Mo K}\alpha) = 0.094 \text{ mm}^{-1}$ ,  $F(000) = 936$ , Reflections collected = 13086, Independent reflections = 3984,  $R(\text{reflections}) = 0.0566$  (2306),  $wR2(\text{reflections}) = 0.1566(3984)$ ,  $S = 0.988$ ,  $N_{\text{par}} = 307$

#### 2.5 Antioxidant assay

To measure antioxidant assays, the concentration of 1,000 mg/L of each plant-derived compound was firstly prepared by dissolving in 95% methanol. A derivative vitamin E with potent antioxidant properties, Trolox, was added as a positive control. The free radical scavenging capacities of the compound was tested *in vitro* by 2,2'-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azino-bis(3-ethyl benzthiazo line-6-sulphonic acid) (ABTS), assay according to the previous method

(21). Briefly, an aliquot (100  $\mu\text{L}$ ) of each sample was carefully mixed with 80  $\mu\text{M}$  DPPH ethanol solution (100  $\mu\text{L}$ ) in a 96-well plate. After incubation in the dark for 30 min at ambient temperature, the absorbance of the solution was measured at 520 nm comparing to an appropriate blank to obtain the DPPH radical (DPPH $^\bullet$ ) bleaching. ABTS radical cation (ABTS $^+$ ) was generated by mixing 2 mM ABTS and 2.45 mM potassium persulfate at a volume ratio of 1:1. The mixture was incubated in the dark at room temperature for 16 h. Then, the absorbance of the solution was adjusted to  $0.70 \pm 0.05$  at 734 nm with ethanol. An aliquot (100  $\mu\text{L}$ ) of each sample was mixed with 100  $\mu\text{L}$  of ABTS $^+$  solution. After 6 min of incubation, the absorbance was then taken at 734 nm.

#### 2.6 Antibacterial activity assay

The antibacterial activity of isolated compounds against *S. aureus* ATCC23235 was evaluated using a modified broth microdilution method from the Clinical and Laboratory Standards Institute (CLSI) (22). Briefly, the bacterial suspension of tested bacteria in Mueller-Hinton broth was mixed with the 2-fold serial dilutions of isolated compounds in 96-well plates. The plates were then incubated at 37  $^\circ\text{C}$  for 24 hours. After incubation, the minimal inhibitory concentration (MIC) of each compound is defined as the lowest concentration of the compounds that completely inhibits the bacterial growth.

#### 3. Results and Discussion

Compound **1** was obtained as a pale yellow powder which was previously reported from the hydrolysis of 1,6-dimethoxy-5-toluene-*p*-sulphonyloxyxanthone (23). The UV spectrum presented absorption bands characteristic of xanthone at 240, 263 and 344 nm. The IR spectrum also presented the main peaks of xanthone such as the hydroxyl group at  $3280 \text{ cm}^{-1}$ , the conjugated carbonyl group at  $1650 \text{ cm}^{-1}$  and an aromatic at  $1575 \text{ cm}^{-1}$  (9). The  $^1\text{H}$  NMR spectrum of **1** showed AMX pattern signals of aromatic protons at  $\delta_{\text{H}} 6.94$  (d,  $J = 8.0 \text{ Hz}$ ), 7.67 (t,  $J = 8.0 \text{ Hz}$ ) and 7.11 (d,  $J = 8.0 \text{ Hz}$ ), signals of two *ortho*-coupled aromatic protons at  $\delta_{\text{H}} 7.10$  (d,  $J = 8.7 \text{ Hz}$ ) and 7.67 (d,  $J = 8.7 \text{ Hz}$ ) and singlet signals of methoxyl protons at  $\delta_{\text{H}} 3.94$  (s, 1- $\text{OCH}_3$ ) and  $\delta_{\text{H}} 3.98$  (s, 6- $\text{OCH}_3$ ). The  $^{13}\text{C}$  NMR data of **1** presented 15 carbon signals, including a conjugated ketone carbonyl carbon ( $\delta_{\text{C}} 174.3$ ), seven quaternary aromatic carbons ( $\delta_{\text{C}} 160.9$ , 158.2, 151.6, 144.3, 134.0, 117.9 and 112.0), five methine aromatic carbons ( $\delta_{\text{C}} 134.6$ , 116.4, 109.7, 108.2, 106.0) and two methoxy carbons ( $\delta_{\text{C}} 55.9$  and 55.6). In the HMBC spectrum of compound **1**, the correlations between H-3 and C-1 ( $\delta_{\text{C}} 160.9$ ) and H-8 and C-6 ( $\delta_{\text{C}} 151.6$ ) and the NOESY correlations H-2 $\leftrightarrow$ 1- $\text{OCH}_3$  and H-7 $\leftrightarrow$ 6- $\text{OCH}_3$  (indicated the location of the two methoxy groups at C-1 and C-6,

respectively (Figure 2). The significantly high field-shifted signal at  $\delta_c$  134.0 as well as the HMBC correlation from H-7 to C-5 ( $\delta_c$  134.0) indicated that the hydroxyl group was attached to C-5 (Figure 2). Thus, compound **1** was assigned as 5-hydroxy-1,6-dimethoxyxanthone.

Compound **2** was previously isolated from *Swertia mileensis* (24) and *Swertia franchetiana* (25). In this study, it was isolated as a pale yellow solid. Compound **2** presented similar UV and IR data to **1**, indicating that it was also a xanthone derivative. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of **2** were also similar to those of **1**. The differences were that, instead of the signals of *ortho*-coupled and AMX aromatic protons produced by **1**, compound **2** presented one singlet signal of an aromatic proton at  $\delta_H$  7.13 (H-8,  $\delta_c$  94.7)

and signals of *meta*-coupled aromatic protons at  $\delta_H$  6.48 (d,  $J = 2$  Hz, H-2,  $\delta_c$  94.8) and  $\delta_H$  6.74 (d,  $J = 2$  Hz, H-4,  $\delta_c$  92.7). Furthermore, the  $^1\text{H}$  NMR spectrum of **2** exhibited an additional two methoxy proton signals at  $\delta_H$  3.91 and 3.94. These methoxy groups were placed at C-7 and C-3, respectively, due to the NOESY correlations (H-4 $\leftrightarrow$ 3-OCH<sub>3</sub> ( $\delta_H$  3.91), H-8 $\leftrightarrow$ 7-OCH<sub>3</sub> ( $\delta_H$  3.94)) as well as the carbon chemical shifts in the dioxysubstituent aromatic ring of compound **2** at C-1 ( $\delta_c$  161.7), C-2 ( $\delta_c$  94.8), C-3 ( $\delta_c$  165.3) and C-4 ( $\delta_c$  92.7) corresponded to those for 1,3-dimethoxyxanthone at  $\delta_c$  161.7, 94.8, 164.6 and 92.5, respectively (26) (Figure 2). Therefore, the structure of **2** was determined to be 5-hydroxy-1,3,6,7-tetramethoxyxanthone.

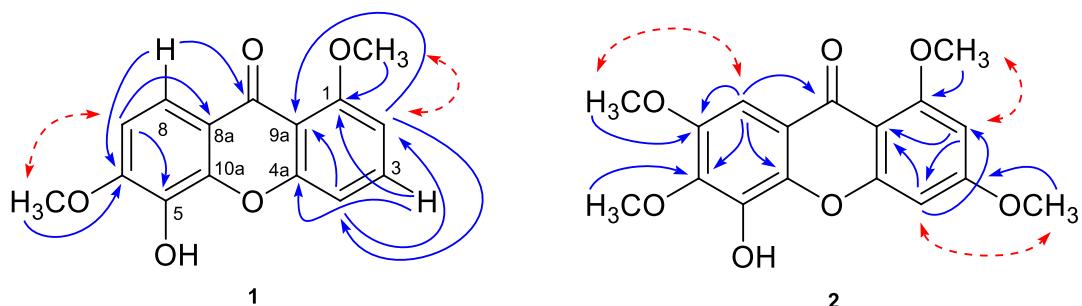


Figure 2 Selected HMBC (blue curved arrows) and NOESY (red dashed arrows) correlations of **1** and **2**.

Mammea A/AA cyclo F, compound **9**, is a 6-acyl-7,8-dihydrofuran derivative of 4-phenyl-5,7-dihydroxycoumarin previously isolated from *Mammea americana* (15) and *M. ferrea* blossoms (14). In this study, the 4-phenylcoumarin, **9**, was isolated as a colourless solid which was recrystallized using EtOAc to yield colourless single crystals. The structure of the crystal was analysed by single-crystal X-ray diffraction (Figure 3).

Compounds **3–5** and **9** were evaluated for their antioxidant activities (Table 1). Xanthone **3** displayed significant antioxidant activity with IC<sub>50</sub> values of 45.0  $\mu\text{g}/\text{mL}$  and 197.0  $\mu\text{g}/\text{mL}$  in the DPPH and ABTS assays, respectively. Compounds **4** and **5** showed inhibitory activity in an ABTS assay with IC<sub>50</sub> values of 264.0  $\mu\text{g}/\text{mL}$  and 212.0  $\mu\text{g}/\text{mL}$ , respectively. All these values were lower than the IC<sub>50</sub> values obtained from the standard antioxidant agent, trolox, in the same assays (77.6 $\pm$ 1.0  $\mu\text{g}/\text{mL}$  in DPPH and 282.9 $\pm$ 1.2  $\mu\text{g}/\text{mL}$  in ABTS). Besides, the xanthone, **4**, and the 4-phenylcoumarin, **9**, showed weak antibacterial activities against *Staphylococcus aureus* ATCC23235 with MIC value of 62.5  $\mu\text{g}/\text{mL}$  and 125.0  $\mu\text{g}/\text{mL}$ , respectively, compared to the antibiotic vancomycin (MIC 0.97  $\mu\text{g}/\text{mL}$ ) (Table 1).

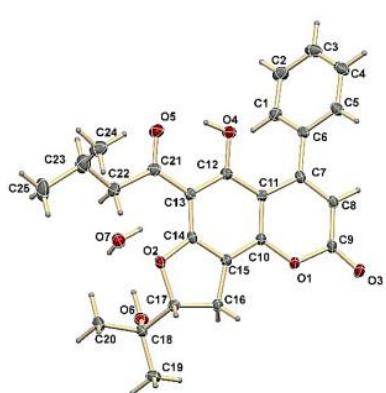


Figure 3 X-ray ORTEP diagram of **9**.

**Table 1** Biological activities of isolated compounds.

Compounds	Radical scavenging properties		Antibacterial activity <i>S. aureus</i> ATCC 23235
	IC <sub>50</sub> (µg/mL) DPPH	IC <sub>50</sub> (µg/mL) ABTS	
<b>3:</b> 1,5,6-trihydroxyxanthone	45.0±0.001	197.0±0.002	>1,000
<b>4:</b> 1,3,7-trihydroxyxanthone	8513±0.350	264.0±0.002	62.5
<b>5:</b> 1,7-dihydroxy-3-methoxyxanthone	8903±0.890	212.0±0.002	>1,000
<b>9:</b> mammea A/AA cyclo F	-	-	125.0
troloxa <sup>a</sup>	77.6±1.06	282.9±1.29	
vancomycin <sub>a</sub>			0.97

<sup>a</sup> Trolox and vancomycin were used as the standard drugs.

#### 4. Conclusions

In summary, five xanthones and four phenylcoumarins were isolated from twigs of *M. ferrea* and structure elucidated. The <sup>1</sup>H and <sup>13</sup>C-NMR spectra of compounds **1** and **2** were completely assigned since there are no NMR data that had been previously published from these compounds. The antioxidant properties of compounds **3–5** and **9** were assessed using DPPH and ABTS scavenging capacity assays. Compound **3** was found to be more active than the other compounds with IC<sub>50</sub> values of 45.0 µg/mL and 197.0 µg/mL in DPPH and ABTS assays, respectively. Compounds **4–7** were isolated from this plant for the first time. The structure of 4-phenylcoumarin **9** was analysed by single-crystal X-ray diffraction.

#### Acknowledgements

The research was sponsored by the Graduate School, Prince of Songkla University and TRF Senior Research Scholar (Grant No. RTA6180006), the Thailand Research Fund. A.C. thanks the Natural Product Research Center of Excellence, Prince of Songkla University and Suratthani Rajabhat University SRU for partial financial support. The authors would also like to thank Mr. Thomas Duncan Coyne for his assistance with the English.

#### Declaration of conflicting interests

The authors declared that they have no conflicts of interest in the research, authorship, and this article's publication.

#### References

- Chanda S, Rakholiya K, Parekh J. Indian medicinal herb :Antimicrobial efficacy of *Mesua ferrea* L. seed extracted in different solvents against infection causing pathogenic strains. *J. Acute Dis.* 2013; 2: 277–81.
- Manandhar NP. Medico botany of Gorkha district, Nepal - An elucidation of medicinal plants. *Int. J. Crude Drug Res.* 1990; 28: 17–25.
- Lim TK. Edible medicinal and non-medicinal plant. New York: Springer; 2012.
- Chakraborty T, Das MK. Oil of *Mesua ferrea* L. seed as a promising pharmaceutical excipient in lipid based nanoformulation. *J. Appl. Pharm. Sci.* 2017; 7: 133–41.
- Chahar MK, Sanjaya Kumar DS, Lokesh T, Manohara KP. In-vivo antioxidant and immunomodulatory activity of mesuol isolated from *Mesua ferrea* L. seed oil. *Int. Immunopharmacol.* 2012; 13: 386–91.
- Rahman SMM, Shabnom S, Quader MA, Hossain MA .Phytochemical study on the ethyl acetate extract of the leaves of *Mesua ferrea* Linn. *Indones. J. Chem.* 2008; 8 :242–44.
- Wetwitayaklung P, Phaechamud T, Limmatvapirat C, Keokitichai S. The study of antioxidant activities of edible flower extracts. *Acta Hortic.* 2008;786:185–92.
- Keawsa-ard S, Kongtaweeert S. Antioxidant, antibacterial, anticancer activities and chemical constituents of the essential oil from *Mesua ferrea* leaves. *Chiang Mai J. Sci.* 2012; 39: 455–63.
- Chukaew A, Saithong S, Chusri S, Limsuwan S, Watanapokasin R, Voravuthikunchai SP, Chakthong S. Cytotoxic xanthones from the roots of *Mesua ferrea* L. *Phytochemistry.* 2019; 157: 64–70.
- Aihua G, Jun L, Hongzheng F, Wenhan L. Xanthone derivatives from medicinal plant *Swertia mileensis..* *Zhongcaoyao.* 2003; 34: 107-9.
- Singh S, Gray AI, Waterman PG. Mesuabixanthone-A and mesuabixan thone-B : bis-xanthones from the stem bark of *Mesua ferrea* (Guttifera). *Nat. Prod. Lett.* 1993; 3: 53–8 .
- Atkinson JE, Gupta P, Lewis JR. Some phenolic constituents of *Gentiana lutea*. *Tetrahedron.* 1969; 25: 1507–11.
- Guillet D, Hélesbeux JJ, Séraphin D, Sévenet T, Richomme P, Bruneton J. Novel cytotoxic 4-phenylfuran coumarins from *Calophyllum dispar*. *J. Nat. Prod.* 2001; 64: 563–68.
- Verotta L, Lovaglio E, Vidari G, Finzi PV, Neri MG, Raimondi A, Parapini S, Taramelli D, Riva A, Bombardelli E. 4-Alkyl- and 4-phenylcoumarins from *Mesua ferrea* as promising multidrug resistant antibacterials. *Phytochemistry.* 2004; 65: 2867–79.

15. Crombie L, Games DE, McCormick A. Extractives of *Mammea americana* L. Part II. The 4-phenylcoumarins. Isolation and structure of Mammea A/AA, A/A cyclo D, A/BA, A/AB, and A/BB. *J. Chem. Soc. (C)*. 1967; 2553–59.
16. Bandaranayake WM, Selliah SS, Sultanbawa MUS, Games DE. Xanthones and 4-phenylcoumarins of *Mesua thwaitesii*. *Phytochemistry*. 1975; 14: 265–9.
17. Bruker. SMART, SAINT and SADABS. Bruker AXS Inc.; Madison: Wisconsin, USA; 2003.
18. Farrugia LJ. WinGX and ORTEP for Windows: an update. *J. Appl. Crystallogr.* 2012; 45: 849–54.
19. Macrae CF, Bruno IJ, Chisholm JA, Edgington PR, McCabe P, Pidcock E, Rodriguez-Monge L, Van de Streek J, Taylor R, Wood PA. Mercury CSD 2.0 - new features for the visualization and investigation of crystal structures. *J. Appl. Crystallogr.* 2008; 41: 466–70.
20. Sheldrick GM. Crystal structure refinement with SHELXL. *Acta Crystallogr. C*. 2015; C71: 3–8.
21. Saeed N, Khan MR, Shabbir M. Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complement. Altern. Med.* 2012; 12: 1–12.
22. Clinical and Laboratory Standards Institute: CLSI. M07-A8-Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; Approved Standard-Eighth Edition. Clinical and Laboratory Standards Institute. Wayne, Pennsylvania, USA; 2009.
23. Jackson B, Locksley HD, Scheinmann F. Extractives from Guttiferae. Part V. Scriblitifolic acid, a new xanthone from *Calophyllum scriblitifolium* Henderson and Wyatt-Smith. *J. Chem. Soc. (C)*. 1967: 785–96.
24. Guo A, Li J, Fu H, Lin W. Xanthone derivatives from medicinal plant *Swertia mileensis*. *Zhongcaoyao*. 2003; 34: 107–9.
25. Wang S, Xiao H, Liu X, Du Y, Han X, Ling X. Study on xanthone components of *Swertia franchetiana*. *Zhongcaoyao*. 2003; 34: 878–79.
26. Chaudhuri RK, Zymalkowski F, Frahm AW.  $^{13}\text{C}$  NMR-spectroscopy of polymethoxyxanthones. *Tetrahedron*. 1978; 34: 1837–40.