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Research Article

# X-ray phase technology shed new light on Weberian apparatus evolution: context and aim of the case study of †*Chanoides* (Otophysi, *incertae sedis*)

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**Abstract** - Otophysan fish are identified as a natural group and have been studied as such since the first half of the 19th century. This very speciose group largely dominates the vertebrate diversity of inland waters today. In the 1980's passionate debates concerned the otophysan radiation and notably their primary habitat: are they primitively freshwaters or marine animals? To answer we need to better understand the phylogenetic relationships including those of the fossils and thus we need to enhance our knowledge on key bony structures such as the Weberian Apparatus. In this context we propose a CT scan approach, for a specimen of †*Chanoides macropoma*. In the last years, palaeontology benefited from the developments of phase contrast X-ray microtomography. We expect great advances to investigate the Weberian apparatus and its evolution in Otophysan fishes.

**Keywords:** Weberian apparatus, evolution, stem otophysan, fossil

## 1. Introduction

Otophysan fish are identified as a natural group and have been studied as such since the first half of the 19th century. This very speciose group largely dominates the vertebrate diversity of inland waters today. Its diversification starts in the Cretaceous in the fossil record (Mayrinck et al., 2015b; DM and OO current studies) while last fossil calibrated molecular studies give for the clade a Jurassic age, or a Jurassic to Early Cretaceous age, and even an early Cretaceous age when excluding saturated partition for nucleotide substitution (respectively Near et al., 2012, Broughton et al., 2013, Dornburg et al., 2014). The success of these fishes in continental freshwaters is generally imputed to the great development in their auditory ability in relation with the presence of an original structure called Weberian Apparatus. Since the first definition of the group, Sagemehl (1885) recognised Ostariophysi (= Otophysi according to current taxonomy) based on the presence of this unique morphological and osteological feature. First described by Ernst Heinrich Weber (Weber, 1820), it consists in a deep modification of at least the first four vertebrae and associated bones and tissues and allows the transmission and amplification of pressure variation from the swimbladder to the inner ear. It includes a series of small paired bony elements (claustrum, scaphium, intercalarium, tripus) linked by the interossicular ligament (e.g. Rosen and Greenwood, 1970). Its presence is now recognised as the apomorphy of the clade Otophysi and lacks at such in the sister ostariophysan group Gonorynchiformes (e.g. Rosen and Greenwood, 1970; Grande and de Pinna, 2004). Moreover, the Weberian structures present a wide diversity in their relative dimension, shape and organisation which allow distinguishing the members of the five otophysan orders, i.e. Characiformes (including piranhas, tiger fish and tetras), Cypriniformes (including carps, barbs and minnows), Siluriformes (catfishes) and Gymnotiformes (electric eels). Otophysan fish are mostly freshwaters and include most of the specific diversity of the Ostariophysi while Gonorhynchiformes (including milkfish and shellears) is a marine and comparatively depauperate clade.

In the 1980's passionate debates concerned the otophysan radiation and notably their primary habitat: are they primitively freshwaters or marine animals? How to explain that Siluriformes and Characiformes originate onto Western Gondwana dependences (South America and/ or Africa) while Cypriniformes are primitively Laurasian animals? To resume in the main lines, several marine fossil fish were described as stem otophysan which pleads for a marine and late Mesozoic origin of the clade. Conversely, a freshwater origin is assumed based on their modern dominant ecology and supports an earlier origin on Pangea at the very beginning of the Mesozoic, or later rafting between the landmasses of the dislocating Pangea. The different scenarios for their diversification oppose vicariance and dispersion models (see for instance Briggs, 2005; Otero et al., 2008 for details). In the background, there were also debates about the phylogenetic position of demonstrated (Davesne et al., 2016).

extant and fossil taxa compared to major clades, and even about the relevance of fossils to reconstruct evolutionary process due either to a low confidence in morphological phylogenies or in the partial information yielded by fossils. We are convinced that fossils are relevant to retrace early stages of ancient clade diversification, and that dedicated studies may even allow to resolve remaining discrepancies between molecular and morphological studies as recently

Progress in the reliability of phylogenetic assumption based on both morphological (e.g. Fink and Fink, 1981, 1996) and molecular data (Calcagnotto et al., 2005; Peng et al., 2006; Sullivan et al., 2006; Saitoh et al., 2011; Chen et al., 2013) and progress in understanding developmental aspects of some ostariophysan phylogenetic key bony features (Coburn and Futey, 1996; Bird and Mabee, 2003; Britz and Hoffman, 2006; Hoffman and Britz, 2006; Britz and Moritz, 2007) led us (DM and OO) to undertake a review of fossil ostariophysan fish (Mayrinck et al., 2015a, b, in review). The final aim is to be able to include confidently fossil data in comprehensive reconstruction of the ostariophysan diversification. So far we were able to confirm a probable stem-otophysan status solely to †Chanoides macropoma, a fossil fish from marine Middle Eocene deposits of the Monte Bolca (Mayrinck et al., 2015a, b, in review). The detailed anatomical studies of its Weberian apparatus by Patterson (1984) and by Mayrinck et al. (2015b) lead to conclude that †Chanoides macropoma presents certainly an Otophysan anatomy but certainly not one of the modern patterns displayed by the members of

In that frame, investigating as far as possible the Weberian Apparatus of †*Chanoides macropoma* appears critical, and was the subject of a dedicated 3D-imaging study based on one of the most powerful X-ray microtomography available today, i.e. the Synchrotron in Grenoble (ESRF). Methodological details and preliminary results on the study of the fossil in propagation-based phase-contrast X-ray microtomography (PBPCT) are given below in the paper.

#### 2. Material and methods

The specimen NHM P. 51216, housed in the Natural History Museum, London was imaging in PBPCT. It has been prepared by transfer method (Toombs and Rison, 1959) and was studied by Patterson (1984) and Mayrinck *et al.* (2015b). They provided a detailed and mainly convergent description of the species and notably of its Weberian Apparatus, including information on NHM P. 51216.

# 2.1. Microtomography acquisition, reconstruction and vizualisation

The microtomogram was obtained by propagation-based phase-contrast X-ray synchrotron microtomography at the ID19 beamline of the European Synchrotron Radiation Facility (Boistel *et al.*, 2001; Tafforeau *et al.*, 2006; Sutton, 2008; Zanette *et al.*, 2013). The energy of pink beam was tuned to 41 keV to obtain a suitable signal/noise ratio with this kind of sample. The X-ray detector was an in-house FReLoN (Fast Readout Low Noise) camera (Labiche *et al.*, 2007). Images of 5 levels were taken in the phase contrast mode with pixels size of 7.46  $\mu$ m. For the tomographic reconstruction, a phase retrieval approach was used from a single long propagation distance using the Paganin

filter of the PyHST2 algorithm (Paganin *et al.*, 2002; Mirone *et al.*, 2014). Finally, volume renderings were obtained after semi-automatic segmentation of the volumes of interest and iso-surfaces were produced using Avizo 7.1 (VSG, Visualization Sciences Group).

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# 2.2. Natural light observation and photographs

The specimen was observed with a binocular (Leica MZ 75) and photographs were realised with a grazing light with a NIKON Coolpix S9900.

#### 3. Results

# 3.1. Observation of the Weberian apparatus of NHM P. 51216 observed in natural light

The specimen is probably the best available Weberian apparatus for †*Chanoides macropoma* (Fig. 1A). Most of the information presented for the species in the papers of Patterson (1984) and Mayrinck *et al.* (2015b) is easily recovered. In brief we were able to recognise: the claustrum, scaphium and intercalarium; the third centrum (which is the shortest); the small process which is interpreted as a tripus by Patterson (1984) but not by Grande and Pinna (2004) and Mayrinck *et al.* (2015b); an os suspensorium that projects backward; the contiguous supraneurals 2 and 3 (Fig. 1).

# 3.2. Volume rendering of Weberian area in NHM P. 51216

On an iso-surface of the Weberian area obtained by a threshold (Fig. 2) we recognize the presence of the Weberian apparatus. Further segmentation will allow isolating the various Weberian elements and produce a 3D mesh. We will also be able to discuss further the modern and archaic aspects of its anatomy when compared with the Weberian apparatus displayed by the members of the four extant otophysan orders.



**Fig. 1.** Detail photograph of the Weberian apparatus of †*Chanoides macropoma* NHM P. 51216 in natural light. Scale bar is 5 mm.



**Fig. 2.** Volume rendering of the Weberian apparatus of †*Chanoides macropoma* NHM P. 51216.

#### 4. Conclusive Discussion

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In the last years, palaeontology benefited from the developments of phase contrast X-ray microtomography. For instance, the methodologies developed at the ID 19 at the ESRF shed new light on evolution through the study of emblematic fossils (e.g. http://paleo.esrf.eu). We expect great advances to investigate the Weberian Apparatus and its evolution in Otophysan fishes.

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# References

- Bird, N. C. and Mabee, P. M. 2003. Developmental morphology of the axial skeleton of the zebrafish, *Danio rerio* (Ostariophysi: Cyprinidae). Developmental Dynamics 228, 337–357.
- Boistel, R., Swoger, J., Kržič, U., Fernandez, V., Gillet, B. and Reynaud, E. G. 2011. The future of three dimentional microscopic imaging in marine biology. Marine Ecology 32(4), 438-452.
- Briggs, J. C. 2005. The biogeography of otophysan fishes (Ostariophysi, Otophysi): a new appraisal. Journal of Biogeography 35, 287–294.
- Britz, R. and Hoffman, M. 2006. Ontogeny and homology of the claustra in otophysan Ostariophysi (Teleostei). Journal of Morphology 267, 909–923.
- Britz, R. and Moritz, T. 2007. Reinvestigation of the osteology of the miniature African freshwater fishes

- Cromeria and Grasseichthys (Teleostei, Gonorynchiformes, Kneriidae), with comments on Kneriidae relationships. Mitteilungen aus dem Museum für Naturkunde in Berlin, Zoologische Reihe 83 (1), 3–42.
- Broughton, R. E., Betancur-R., R., Li, C., Arratia, G. and Ortí, G. 2013. Multi-locus phylogenetic analysis reveals the pattern and tempo of bony fish evolution. PLOS Currents Tree of Life. Edition 1. doi:10.1371/currents.tol.2ca8041495ffafd0c92756e75247483e.
- Calcagnotto, D., Schaefer, S. A. and DeSalle, R. 2005. Relationships among characiform fishes inferred from analysis of nuclear and mitochondrial gene sequences. *Molecular* Phylogenetics and Evolution 36, 135–153
- Chen, W-J., Lavoué, S. and Mayden, R., 2013. Evolutionary origin and early biogeography of otophysan fishes (Ostariophyi: Teleostei). Evolution 67-8, 2218–2239.
- Coburn, M. M. and Futey, L. M. 1996. The ontogeny of supraneurals and neural arches in the cypriniforms Weberian Apparatus (Teleostei: Ostariophysi). Zoological Journal of the Linnean Society 116, 333–346.
- Davesne, D., Gallut, C., Barriel, V., Lecointre, G., Janvier, P. and Otero, O. 2016. Building a consensus for deep intrarelationships of spiny-rayed teleosts (Acanthomorpha) with a dedicated sampling including fossil taxa. Frontiers in Ecology and Evolution 4, 129, pp. 20.
- Dornburg, A., Townsend, J. P., Friedman, M. and Near, T. J. 2014. Phylogenetic informativeness reconciles ray-finned fish molecular divergence times. BMC Evolutionary Biology 16, pp. 14.
- Fink, S.V. and Fink, W. L. 1981. Interrelationships of the ostariophysan fishes (Teleostei). Zoological Journal of the Linnean Society 72 (4), 297–353.
- Fink, S. V. and Fink, W. L. 1996. Interrelationships of Ostariophysan Fishes (Teleostei). In: Stiassny, M. Parenti, L. and Johnson D. (Eds.), Interrelationships of Fishes. Academic Press, San Diego, pp. 209–249.
- Grande, T. and de Pinna, M. 2004. The evolution of the Weberian Apparatus: A phylogenetic perspective. In: Arratia, G. and Tintori, A. (Eds.), Mesozoic fishes 3: Systematics, Paleoenvironments, and Biodiversity. Verlag Dr. Friedrich Pfeil, München, pp. 429–448
- Hoffmann, M. and Britz, R. 2006. Ontogeny and homology of the neural complex of otophysan Ostariophysi. Zoological Journal of the Linnean Society 175, 301–330.
- Labiche, J. C., Mathon, O., Pascarelli, S., Newton, M.A., Ferre, G. G., Curfs, C, Vaughan, G., and Homs A. 2007. The fast readout low noise camera as a versatile x-ray detector for time resolved dispersive extended x-ray absorption fine structure and diffraction studies of dynamic problems in materials science, chemistry, and catalysis. Review of Scientific Instruments 78(9), 091301–091311.
- Mayrinck D., Brito, P. M. and Otero, O. 2015a. Anatomical review of † Salminops ibericus, a Teleostei incertae

- sedis from the Cenomanian of Portugal, anciently assigned to Characiformes and possibly related to crossognathiform fishes. Cretaceous Research 56, 66–75.
- Mayrinck, D., Brito P. M. and Otero, O. 2015b. Review of the osteology of the fossil fish formerly attributed to the genus †*Chanoides*, systematic and implications for the definition of otophysan bony characters. Journal of Systematic Palaeontology 13(5), 397–420.
- Mayrinck, de D., Brito, P. M., Meunier, F.J., Alvarado-Ortega, J. and Otero, O. In review. †Sorbinicharax verraesi: an unexpected case of a benthic fish outside Acanthomorpha in the Upper Cretaceous of the Tethysian Sea.
- Mirone, A., Brun, E., Gouillard, E., Tafforeau, P. and Kieffer, J. 2014. Pyhst2: a hybrid distributed code for high speed tomographic reconstruction with iterative reconstruction and a priori knowledge capabilities. Nuclear Instruments and Methods and Physics Research Section B: Beam interactions with Material and Atoms 324, 41–48.
- Near, T. J., Dornburg, A., Eytan, R. I., Keck, B. P., Smith, W. L., Kuhn, K. L., Moore, J. A., Price, S. A., Burbrink, F. T., Friedman, M., and Wainwright, P. C. 2012. Phylogeny and tempo of diversification in the superradiation of spiny-rayed fishes. Proceedings of the National Academy of Sciences 110, 12738–12743.
- Otero, O., Valentin, X. and Garcia, G. 2008. Cretaceous characiform fishes (Teleostei: Ostariophysi) from Northern Tethys: description of new material from the Maastrichtian of Provence (Southern France) and palaeobiogeographical implications. In: L. Cavin, A. Longbottom and M. Richter (Eds.), Fishes and the Break-up of Pangea. Geological Society, Special Publications, London, pp. 155–164.
- Paganin, D., Mayo, S. C., Gureyev, T. E., Miller, P. R. and Wilkins, S. W. 2002. Simultaneous phase and amplitude extraction from a single defocused image of a homogeneous object. Journal of microscopy 206(1), 33–40.
- Patterson, C. 1984. *Chanoides*, a marine Eocene otophysan fish (Teleostei: Ostariophysi). Journal of Vertebrate Palaeontology 4 (3), 430–456.
- Peng, Z., He, S., Wang, J., Wang, W. and Diogo, R. 2006. Mitochondrial molecular clocks and the origin of the major Otocephalan clades (Pisces: Teleostei): A new insight. Gene 360, 113–124.
- Rosen, D. E. and Greenwood, P. H. 1970. Origin of the Weberian apparatus and the relationships of the ostariophysan and gonorynchiform fishes. American Museum Novitates 2428, 1–25.
- Sagemehl, M. 1885. Beiträge zur vergleichenden Anatomie der Fische. III. Das Cranium der Characinidien nebst allgemeinen Bemerkungen über die mit einem Weber'schen Apparat versehenen Physostomenfamilien. Morphologisches Jahrebücher 10, 1–119
- Saitoh, K., Sado, T., Doosey, M. H., Bart. Jr., H. L., Inoue, J. G., Nishida, M., Mayden, R. L. and Miya, M. 2011.

Evidence from mitochondrial genomics supports the lower Mesozoic of South Asia as the time and place of divergence of basal divergence of cypriniform fishes (Actinopterygii: Ostariophysi). Zoological Journal of Linnean Society 161, 633–662.

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- Sullivan, J. P., Lundberg, J. G. and Hardman, M. 2006. A phylogenetic analysis of the major groups of cat-fishes (Teleostei: Siluriformes) using *rag1* and *rag2* nuclear gene sequences. Molecular Phylogenetics and Evolution 41, 636–662.
- Sutton, M. D. 2008. Tomographic techniques for the study of exceptionally preserved fossils. Proceedings of the Royal Society B 275, 1587–1593.
- Tafforeau, P., Boistel, R., Boller, E., Bravin, A., Brunet, M., Chaimanee, Y. and Kay, R. F. 2006. Applications of X-ray synchrotron microtomography for non-destructive 3D studies of paleontological specimens. Applied Physics 83(2), 195–202.
- Toombs, H. and Rixon, A. 1959. The use of acids in the preparation of vertebrate fossils, Department of Paleontology, British Museum.
- Weber E. H. 1820. De Aura et Auditu Hominis et Animalium: pars 1 De Aura Animalium aquatilium. Leipzig, Fleischer G.
- Zanette, I., Daghfous, G., Weitkamp, T., Gillet, B., Adriaens, D., Langer, M., and Baumbach, T. 2013. Looking inside marine organisms with magnetic resonance and X-ray imaging, Imaging marine life 122-184.